HBV Quantitative Real Time PCR Kit
User Manual
For In Vitro Diagnostic Use Only

**Warnings and Precaution**

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**3. Product Description**

Chronic hepatitis B virus (HBV) infection affects over 350 million people worldwide and over 1 million die annually of HBV-related chronic liver disease. Although many individuals eventually achieve a state of nonreplicative infection, the prolonged immunologic response to infection may lead to the development of cirrhosis, liver failure, or hepatocellular carcinoma (HCC) in up to 40% of patients. HBV is an enveloped virus in the family Hepadnaviridae, with partially double-stranded DNA and 42nm in diameter. Its antigenic components includes hepatitis B surface antigen (HBsAg), hepatitis B core antigen (HBcAg), and hepatitis B e antigen (HBeAg). Virus replicates in liver and sheds into blood in high concentration. HBV is transmitted through blood and body fluid.

HBV real time PCR kit contains a specific ready-to-use system for HBV detection (for genotype A~H) through polymerase chain reaction (PCR) in the real-time PCR system. The master contains reagents and enzymes for the specific HBV DNA amplification. Fluorescence is emitted and measured by the fluorescence signal is detected initially (Ct) is proportional to the amount of the specific PCR product. Monitoring the fluorescence intensities during Real Time allows the detection of the accumulating product without having to re-open the reaction tube after the amplification.

**5. Storage**

- All reagents should be stored at -20℃. Storage at +4℃ is not recommended.
- All reagents can be used until the expiration date indicated on the kit label.
- Repeated thawing and freezing (> 3x) should be avoided, as this may reduce the sensitivity of the assay.
- Cool all reagents during the working steps.

**6. Additonally Required Materials and Devices**

- Biological cabinet
- Real time PCR reaction tubes/plates
- Pipets (0.5 μl ~ 1000 μl)
- Disposable gloves, powderless
- Biohazard waste container
- Sterile filter tips for micro pipets
- Desktop microcentrifuge for “eppendorf” type tubes (RCF max. 16,000 x g)

**7. ▶️Warnings and Precaution**

Carefully read this instruction before starting the procedure.

- For in vitro diagnostic use only.
- This assay needs to be carried out by skilled personnel.
- Clinical samples should be regarded as potentially infectious materials and should be prepared in a laminar flow hood.
- This assay needs to be run according to Good Laboratory Practice.
- Do not use the kit after its expiration date.
- Avoid repeated thawing and freezing of the reagents, this may reduce the sensitivity of the test.
- Once the reagents have been thawed, vortex and centrifuge briefly the tubes before use.
- Prepare quickly the Reaction mix on ice or in the cooling block.
- Set up two separate working areas: 1) Isolation of the RNA/ DNA and 2) Amplification/ detection of amplification products.
- Pipets, vials and other working materials should not circulate among working units.
- Use always sterile pipette tips with filters.
- Wear separate coats and gloves in each area.
• Avoid aerosols

8. Sample Collection, Storage and transport
• Collect samples in sterile tubes;
• Specimens can be extracted immediately or frozen at -20°C to -80°C.
• Transportation of clinical specimens must comply with local regulations for the transport of etiologic agents

9. Procedure
9.1 DNA-Extraction
DNA extraction buffer is supplied in the kit, please thaw the buffer thoroughly and spin down briefly in the centrifuge before use. It’s better to use commercial kits for nucleic acid extraction.

1) Take 50μl serum or plasma, add 50μl DNA extraction buffer, and close the tube then vortex for 10 seconds. Spin down briefly in a table centrifuge.
2) Incubate the tube for 10 minutes at 100°C.
3) Centrifuge the tube at 13000rpm for 10 minutes. The supernatant contains the DNA extracted and can be used for PCR template.

Attention:
A. During the incubation, make sure the tube is not open, for the vapor will volatilize into the air and may cause contamination in case the sample is positive.
B. The extraction sample should be used in 3 hours or stored at -20°C for one month.
C. DNA extraction kits are available from various manufacturers. You may use your own extraction systems or the commercial kit based on the yield. For DNA extraction, please comply with manufacturer’s instructions.

9.2 Internal Control
It is necessary to add internal control (IC) in the reaction mix. Internal control (IC) allows the user to determine and control the possibility of PCR inhibition.

9.3 PCR Protocol
The Master Mix volume for each reaction should be pipetted as follows:

1) The volumes of Reaction Mix and Enzyme Mix per reaction multiply with the number of samples, which includes the number of controls, standards, and sample prepared. Molecular Grade Water is used as the negative control. For reasons of unprecise pipetting, always add an extra virtual sample. Mix completely then spin down briefly in a centrifuge.
2) Pipet 36μl (22.5μl for Smart Cycler II) Master Mix with micropipets of sterile filter tips to each Real time PCR reaction plate/tubes. Separately add 4μl (2.5μl for Smart Cycler II) DNA sample, QS1,QS2,QS3,QS4 and negative control to different reaction plate/tubes. Immediately close the plate/tubes to avoid contamination.
3) Spin down briefly in order to collect the Master Mix in the bottom of the reaction tubes.
4) Perform the following protocol in the instrument:
   37°C for 2 min,1 cycle;94°C for 2 min,1 cycle;93°C for 15 sec,60°C for 60 sec,40 cycles.
   Fluorescence is measured at 60°C; FAM and HEX/VIC/JOE channels should be chosen.
5) If you use ABI Prism® system, please choose “none” as passive reference and quencher.

10. Threshold setting: just above the maximum level of molecular grade water.

11. Calibration for quantitative detection: Input each concentration of standard controls at the end of run, and a standard curve will be automatically formed.

12. Quality control:

<table>
<thead>
<tr>
<th>Channel</th>
<th>Ct value</th>
</tr>
</thead>
<tbody>
<tr>
<td>FAM (Target Nucleic Acid)</td>
<td>≥38, and Correlation coefficient of QS curve ≤0.98</td>
</tr>
<tr>
<td>HEX/VIC/JOE (IC)</td>
<td>25–35</td>
</tr>
<tr>
<td>Molecular Grade Water</td>
<td>UNDET</td>
</tr>
<tr>
<td>QS1/QS2/QS3/QS4</td>
<td>≤38</td>
</tr>
</tbody>
</table>

13. Data Analysis and Interpretation
The following results are possible:
1) The Ct value in channel FAM shows ≤38. The result is positive: The sample contains HBV DNA. Quantitative value of samples is automatically reported according to the standard curve.

<table>
<thead>
<tr>
<th>Quantitative Value</th>
<th>Data Analysis and Suggestion</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;5×10^7 IU/ml</td>
<td>HBV DNA Positive; its concentration lower than 5×10^7 IU/ml</td>
</tr>
<tr>
<td>5×10^7~10^8 IU/ml</td>
<td>HBV DNA Positive; the quantitative value for recommendation only</td>
</tr>
<tr>
<td>10^8~10^9 IU/ml</td>
<td>HBV DNA Positive; the quantitative value is valid</td>
</tr>
<tr>
<td>&gt;10^9 IU/ml</td>
<td>1) HBV DNA Positive but the quantitative value for recommendation only</td>
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<tr>
<td></td>
<td>2) Re-test the sample after dilute the sample by several times, making the quantitative value within 10^7~10^8 IU/ml</td>
</tr>
</tbody>
</table>

2) The Ct value in channel FAM shows 38–40, please repeat again. If the result still shows 38–40, it can be considered negative.
3) In channel FAM no signal is detected, at the same time, a HEX/VIC/JOE signal from the Internal Control appears. The sample does not contain any HBV DNA. It can be considered negative.
4) Neither in channel FAM nor in channel HEX/VIC/JOE signal is detected. A diagnostic statement can not be made. Inhibition of the PCR reaction.

For further questions or problems, please contact our technical support at trade@liferiver.com.cn